


Phytophthora identification/detection:
PCR, TaqMan PCR, PCR-RFLP,
SSCP, SSR, AFLP, PLP and DNA
barcodes


Peter Bonants, 1 July 2010
Workshop *Phytophthora* Costa Rica



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Introduction

- Plant Research International
- Wageningen University & Research Centre
- Netherlands
- *Phytophthora*
- Identification & detection: molecular methods
- Quarantine species



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Introduction

- Identification - Detection (techniques)
- Past / Now / Future
- Qualitative or Quantitative, Single or Multiplex?
- Applications for *Phytophthora*

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Introducción

- Identificación y detección de las técnicas
- Pasado, presente y el futuro
- Cualitativa o cuantitativa, individual o multiplex ?
- Aplicaciones de *Phytophthora*
- Por favor, no dude en hacer preguntas si no entiende

Clean Material




What is detection?

- Detection is an activity focused on demonstration of the presence or absence of a certain pathogen, which is suspected to be present in the sample
 - single and multiplex tests
 - specificity, sensitivity
 - diagnostics, monitoring
 - quantitative / qualitative
 - live – dead
 - races – formae speciales

Targets


- Which?
 - Bacteria
 - Viruses
 - Nematodes
 - Fungi
 - Insects
- Where?
 - in plant
 - in water
 - in soil
 - in compost
 - in air



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Techniques for identification/detection:

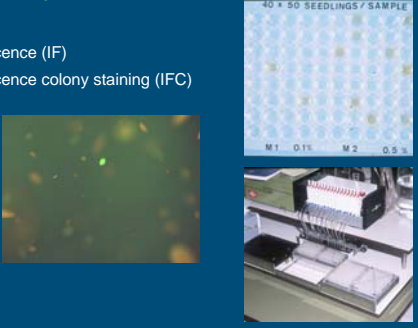
- Based on physiological characteristics
- Based on biological characteristics
- Based on morphological characteristics (microscopy)
- Based on protein/carbohydrate level:
 - Antisera
 - Isozyme-patterns



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Detection with protein based techniques

- ELISA
- Immuno fluorescence (IF)
- Immuno fluorescence colony staining (IFC)
- Luminex system



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Luminex Technology

Color-coded Microspheres

Unique microsphere sets are color-coded using a blend of different fluorescent intensities of two dyes.

100 Color-codes = 100 Simultaneous Tests

Using this method, over 100 distinct microsphere sets can be created.

Microspheres in a Fluid Stream

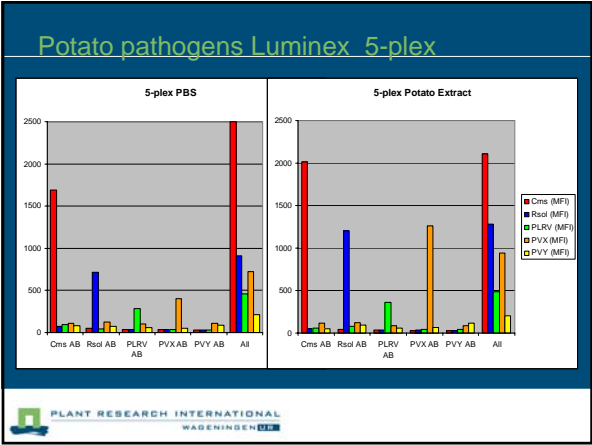
Precision fluidics align the microspheres in single files, and pass them through the beam one at a time.

One Laser Excites Molecular Tags

Reactions are measured with fluorescent intensity and reported in real time.

Second Laser Excites Microsphere

Fluorescent intensity of the microsphere identifies the reporter.



DNA/RNA based techniques

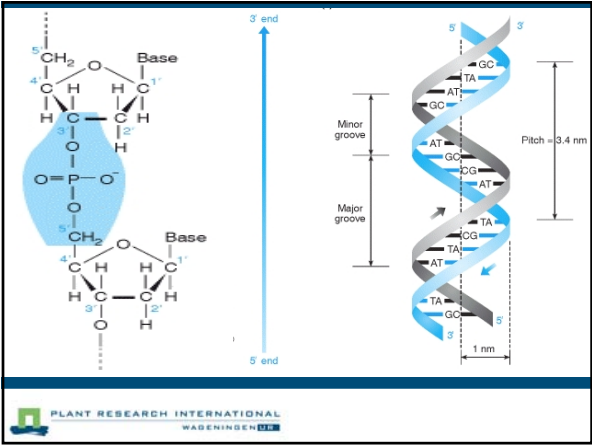
- AFLP
- RAPD-PCR
- SCAR-PCR
- REP-PCR
- ISSR
- SSR
- RFLP
- (RT)-PCR
- DGGE

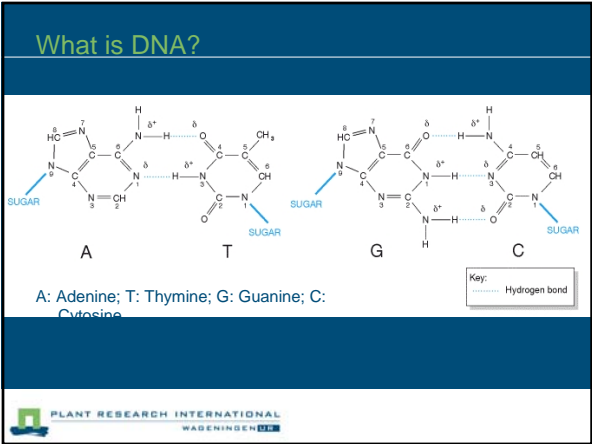
- NASBA
- TaqMan
- Padlock probes

Examples *Phytophthora*

- PCR
- TaqMan PCR
- PCR-RFLP
- AFLP
- RAPD-PCR
- ISSR / SSR
- SSCP
- Padlock probes (PLP)

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Gene sequence analysis

(A) 1 2 3 4 5

(B) Laser Gel Detector Computer Output

TATAAAACATTTTAAAGGTAGTAGCCGATACCTTCTAGTTCGAAAGCCGATGTTGTTGACTATGGTTCAGAAATGGGACCA

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Detection techniques: molecular

- Determine DNA sequence of the perpetrator
- DNA sequence differences between target and non-target
 - often ITS, 16S, 18S, 28S, b-tub, EF1a, Cox1

A CCGAAATCGGACCTTGAGTGC**ACCGTATGCGT**TAGCCTAGTGTACGAGCCCG

B CCGAAATCGGACCTTGAGTGC**AGTACG**TGTTAGCCTAGTGTACGAGCCCGA

C CCGAAATCGGACCTTGAGTGC**AGTAGC**TGTTAGCCTAGTGTACGAGCCCGA

D CCGAAATCGGACCTTGAGTGC**AGATGTTA**CTAGTGTACGAGCCCGA

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Detection techniques: molecular

Blair et al. Fungal Genetics and Biology 45: 266-277 (2008)

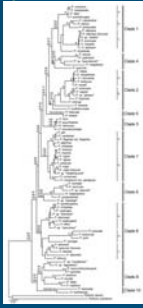
A multi-locus phylogeny for *Phytophthora* utilizing markers derived from complete genome sequences

Jaime E. Blair ^{a,*}, Michael D. Coffey ^{b,*}, Sook-Young Park ^a, David M. Geiser ^a, Seogchan Kang ^a

Genes used:
ITS, B-tubulin, Enolase, Heat Shock Protein 90, 60S Ribosome
Protein L10, LSU rRNA, Cox2, TigA gene fusion, TEF 1a

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Detection techniques: molecular



Blair et al. Fungal Genetics and Biology 45: 266-277 (2008)

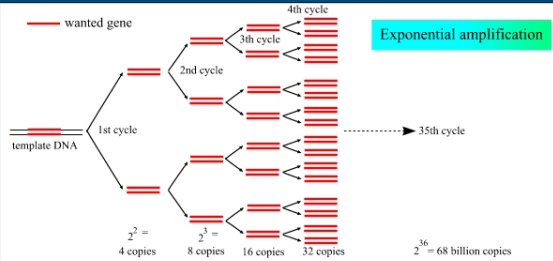
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Detection techniques: molecular



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PCR: polymerase chain reaction



Exponential amplification

template DNA

1st cycle

2nd cycle

3rd cycle

4th cycle

35th cycle

$2^2 = 4$ copies

$2^3 = 8$ copies

$2^4 = 16$ copies

$2^5 = 32$ copies

$2^{36} = 68$ billion copies


(Andy Vriente 1999)

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PCR: Sequence difference perpetrator

Sequence Name	< Pos = 37
- +	AACCCAATTAGTTGGGGG- TCTTGCTGGTG- GCGGCT-
Consensus	40 50 60 70
7 Sequences	
nicotianaITS1.SEQ	AACCCAA- TAGTTGGGGG- TCTTATTTGGCG- GCGGCT-
pseudotsugaITS1.SEQ	AAACCAAATAGTTGGGGG- TCTTGCTGGTG- GCGGCT-
idaeiITS1.SEQ	AAACCAAATAGTTGGGGG- TCTTGCTGGTG- GCGGCT-
oacorumITS1.SEQ	AAACCAAATAGTTGGGGG- TCTTGCTGGTG- GCGGCT-
fragariaeITS1.SEQ	AACCCACTTAGTTGGGGGCTGTCCTG- GCGGCTGGC-
canbivoraITS1.SEQ	AACCCACTTAGTTGGGGGCTAGTCCG- GCGGCTGGC-
cinnamomiITS1.SEQ	AACCCAATTAGTTGGGGGCTGCTCG- GCGGCGGC-

Specific sequence difference for the perpetrator *Phytophthora fragariae*

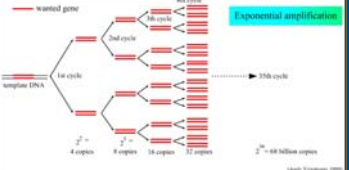
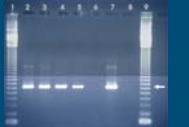




PCR *Phytophthora fragariae*

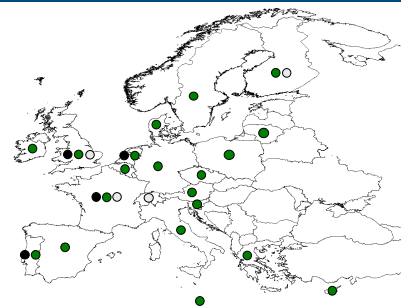
18S ITS-1 5.8S ITS-2 28S

ITS scheme for *P. fragariae*


PCR zoospores *P. fragariae*

TaqMan PCR: EU project Portcheck



Partners



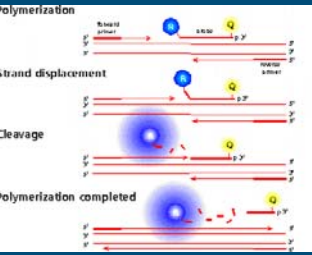
TaqMan PCR

polymerization

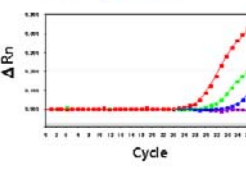
strand displacement

Cleavage

polymerization completed



Real-time monitoring of PCR amplification



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EU-Portcheck: On-site detection

Quality of diagnosis and new diagnostic methods for plant pests: current status and future prospects




lan Barker



- TaqMan PCR protocols
- 10 Q-organisms
- DNA extraction protocols
- On-site system

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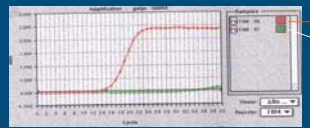
TaqMan PCR *Guignardia citricarpa*



- Symptoms

```
1 TGAATAAGCTTAATCCCTGAAA...GCTTTCGGA...GC G_CITRIGORSA
1 ...T...AC.TC.AT...GTC G_mangiferamay277711
41 AA AAAAGCCGCGGACCTACCTTCA CA...GCTTTCGGA...GC G_CITRIGORSA
39 .C...G...G.A.A.T.G...T...GTC G_mangiferamay277711
```

■ ITS sequentie:



G.
citricarpa
G. *mangiferae*

Specificity TaqMan PCR, non target is *G. mangiferae*

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BioTrove system

3x 3072 PCR reactions in one run; 3x 144 samples

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RAPD PCR *Phytophthora* random amplified polymorphic DNA

PCR with random primers (10-mers)

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Detection techniques: SSCP Single strand conformation polymorphism

<i>P. cactorum</i>	<i>P. cinnamomi</i>	<i>P. nicotianae</i>
SL	SL	SL
15056	1A3	25024
22806	1A10	10011
22807	1B7	113
1008	25102	23809
15022	23801	23800
15024	10112	15025
BL	SL	SL

Fig. 2. Examples of single-patterned species include *P. cactorum*, *P. cinnamomi*, and *P. nicotianae*. The names of representative isolates of individual species are indicated on the top of lanes. SL represents the ssDNA ladder developed in this study.

Ping Kong et al. 2003. Fungal Genetics & Biology 39, 238-249

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Identification techniques: SSCP

Fig. 3. Distinct SSCP patterns of type isolates of 29 species of *Phytophthora* present in a wide mixed. Lane number and SSCP pattern identifier are listed on the top of each lane. Phylogenetic clade numbers of respective species as determined by Cooke et al. (2000) are also indicated for reference. SL is the ssDNA ladder. The numbers on the left of gel indicate the position of major fragments in the ladder.

Ping Kong et al. 2003. *Fungal Genetics & Biology* 39, 238-249

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DNA/RNA based methods

- *Phytophthora ramorum*
- Oak and Rhododendron
- Present in EU and US

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PCR-RFLP *P. ramorum*

restriction fragment length polymorphism

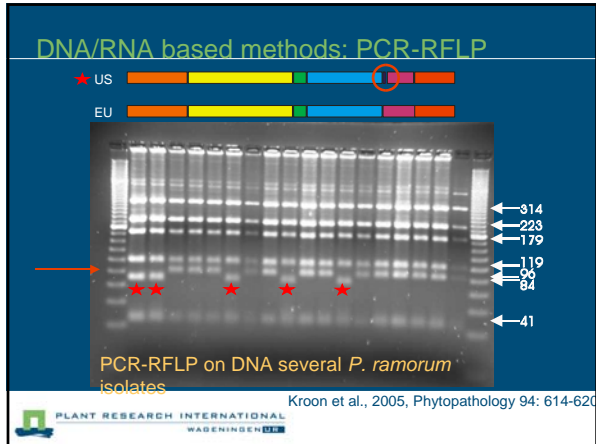
- US
- EU

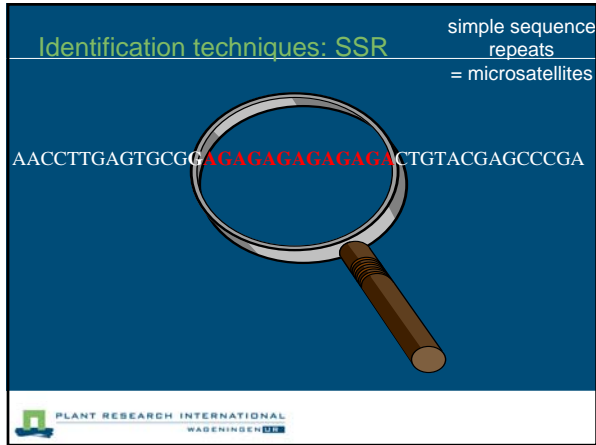
RAATTY Apo I

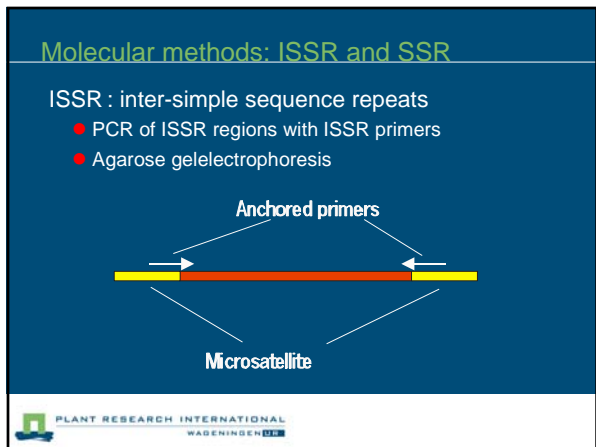
<i>P. ramorum</i> OakVS478	AGGTCAAAT T CATTTTGGTT
<i>P. ramorum</i> OakVS479	AGGTCAAAT T CATTTTGGTT
<i>P. ramorum</i> RhodEU233	AGGTCAAAT C CATTTTGGTT
<i>P. ramorum</i> RhodEU98	AGGTCAAAT C CATTTTGGTT
<i>P. ramorum</i> ViburEU474	AGGTCAAAT C CATTTTGGTT

Gene sequence analysis Cox-1 *P. ramorum*

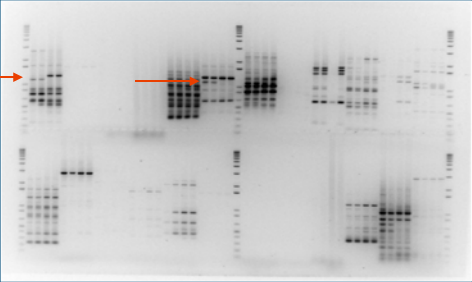
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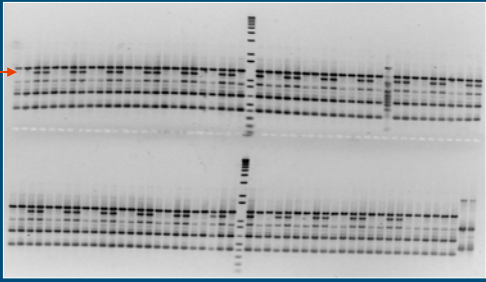
Molecular methods: ISSR *P. ramorum*



ISSR: 2 EU and 2 US isolates with several primers

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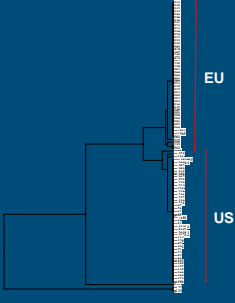
Molecular methods: ISSR *P. ramorum*



ISSR: alternating 2 EU and 2 US isolates with one

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Molecular methods: ISSR *P. ramorum*



ISSR: tree

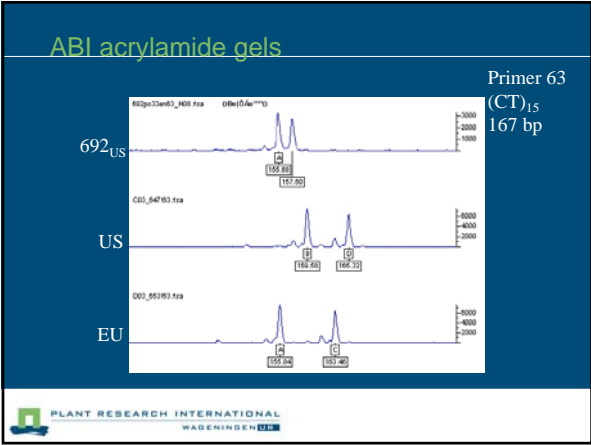
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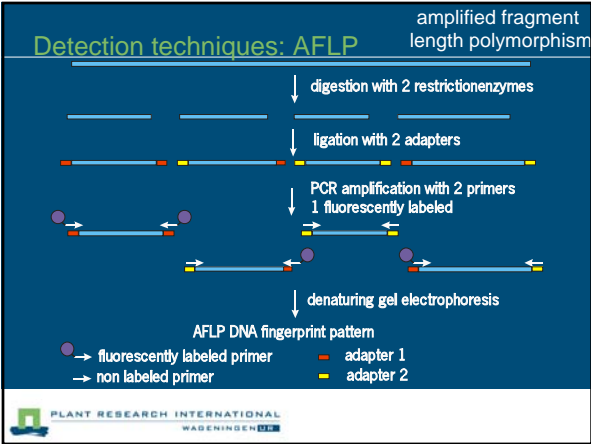
Molecular methods: Microsatellites (SSR)

Microsatellites: SSR (simple sequence repeats), e.g. (AG)_n, (TCG)_n

- PCR of microsatellite region with SSR primers
- Acrylamide gelelectrophoresis

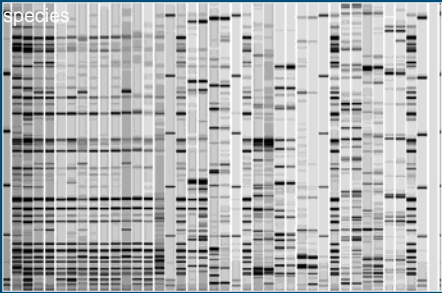
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Molecular methods: AFLP


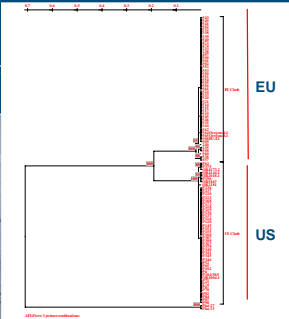
AFLP several *P. ramorum* isolates and other species



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Molecular methods: AFLP

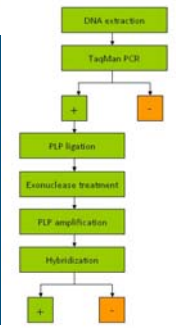

AFLP: 3 primer combinations on many EU and US *P. ramorum* isolates

Ivors et al. 2004. Mycol. Res. 108: 378-392.

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Ouma Padlock-based Universal Multiplex Array

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Universal *Phytophthora* TaqMan PCR

Primer	Sequence	Position
Forward: FITS1_15Ph ITS1	TGC GGA AAG GAT CAT TAC CAC ACC	-17 to +7 of
Reverse: RITS1_279Ph	GCG AGC CTA GAC ATC CAC TG	+11 to +30 of 5.8S rRNA
Probe: all_phy 5.8S rRNA	FAM-TTG CTA TCT AGT TAA AAG CA-MGB	-17 to +3 of

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Kox et al. 2007. Phytopathology 97: 1119-1129.

Padlock Probe Ligation

5'P **T1** **P1** **P2** **Zip code** **T2** OH 3'

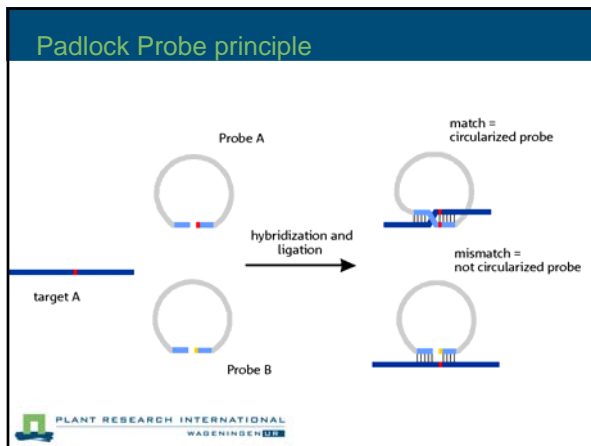
Universal Rev primer Universal Fw primer

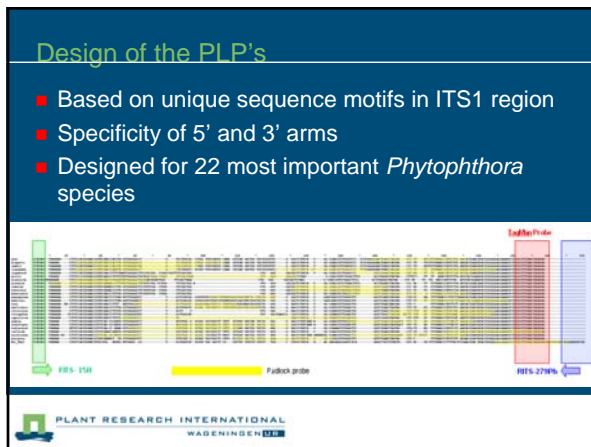
- Circularizing probes
- Long oligonucleotides of ~100bp
- Contain:
 - target complementary regions at both 5' and 3' ends
 - universal primers sites
 - unique sequence identifier (zip-code)

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Padlock Probe principle

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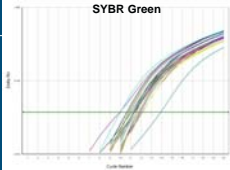



List of designed PLP's

Species	Isolate	Padlock probe
<i>Phytophthora alni</i>	CBS 117376	PLP_alni_50
<i>Phytophthora andina</i>	EC 2425	PLP_andid_162
<i>Phytophthora bostryosa</i>	CBS 533.92	PLP_bostry_193
<i>Phytophthora brassicae</i>	CBS 112277	PLP_bras_78
<i>Phytophthora cactorum</i>	CBS 294.29	PLP_cac_200
<i>Phytophthora cambivora</i>	CBS 114865	PLP_camb_31
<i>Phytophthora capsici</i>	CBS 111333	PLP_caps_150
<i>Phytophthora cinnamomi</i>	CBS 402.48	PLP_cin_30, PLP_cin_46, PLP_cin_55
<i>Phytophthora citricola</i>	CBS 11337	PLP_citri_144
<i>Phytophthora citrophthora</i>	CBS 111338	PLP_citro_154*
<i>Phytophthora cryptogea</i>	CBS 307.62	PLP_cryp_73
<i>Phytophthora fragariae</i>	CBS 309.62	PLP_frag_NAR, PLP_frag_28
<i>Phytophthora hibernalis</i>	CBS 119904	PLP_hybn_96
<i>Phytophthora humicola</i>	CBS 114082	PLP_humici_129*
<i>Phytophthora infestans</i>	PI-98189	PLP_infes_161
<i>Phytophthora lateralis</i>	CBS 102608	PLP_lal_93
<i>Phytophthora megasperma</i>	CBS 118733	PLP_megasp_105
<i>Phytophthora multivesiculata</i>	CBS 101593	PLP_multives_232
<i>Phytophthora nicotianae</i>	CBS 304.29	PLP_nic_166, PLP_nic_186
<i>Phytophthora portii</i>	CBS 181.87	PLP_por_100
<i>Phytophthora primulae</i>	CBS 275.74	PLP_prim2_77
<i>Phytophthora ramorum</i>	CBS 101553	PLP_ram_94
<i>Phytophthora rosacearum</i>	CBS 117690	PLP_rosa_136
<i>Phytophthora tenaculata</i>	CBS 412.96	PLP_tentac_171

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PLP amplification

Primer P1-f19-Biot
P2-r20-Phos

Sequence
5' biotin - CGA GAT GTA CCG CTA TCG T
5' phosphate - TCA TGC TGC TAA CCG TCG AG

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PLP based target signature analysis

Hybridization device



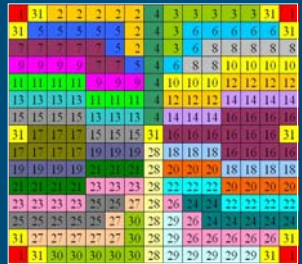

Universal Microarray

- cZipCode oligo's spotted on epoxy-coated slides
- PLP with ZipCode will hybridize on array
- ZipCodes ensure universal hybridization conditions
- ZipCodes provide flexible detection

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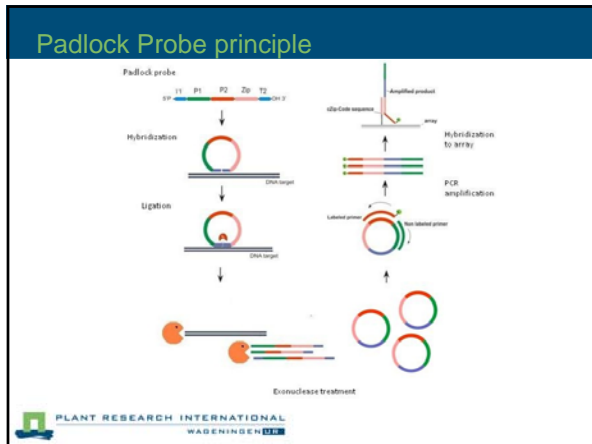
PLP based target signature analysis

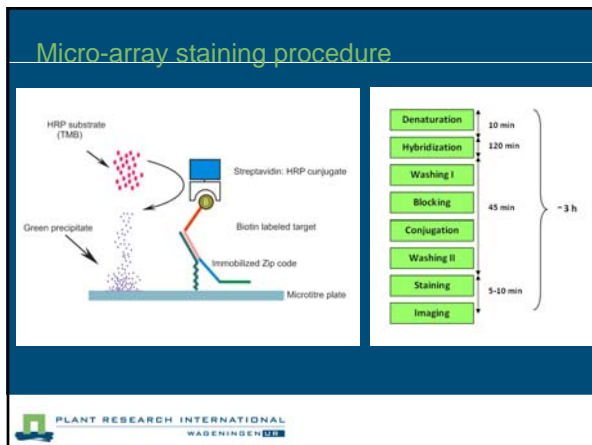
Micro-array design

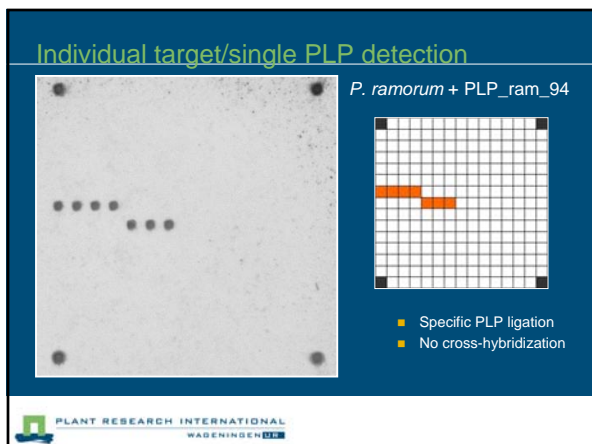


1	Corner Spot	17	PLP_prim2_77
2	PLP_citr6_154	18	PLP_bres_78
3	PLP_boty_193	19	PLP_per_100
4	PLP_nic_166	20	PLP_cin_56
5	PLP_tentac_171	21	PLP_cin_46
6	PLP_andid_162	22	PLP_cin_30
7	PLP_inke6_181	23	PLP_cimb_31
8	PLP_cin_144	24	PLP_cin_50
9	PLP_multives_232	25	PLP_cyp_73
10	PLP_rosa_136	26	PLP_frag_28
11	PLP_humid_129	27	PLP_frag_NAR
12	PLP_megasp_105	28	PLP_nic_186
13	PLP_hybn_96	29	PLP_caps_150
14	PLP_lut_83	30	PLP_cps_200
15	PLP_ran_94	31	PLP_All_Phyt_2004
		32	CH2 (Immobil control)

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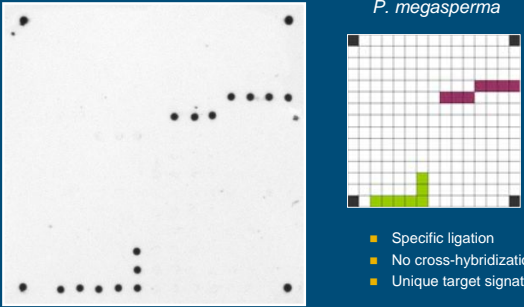






Individual target detection with a mixture of PLP's

P. megasperma

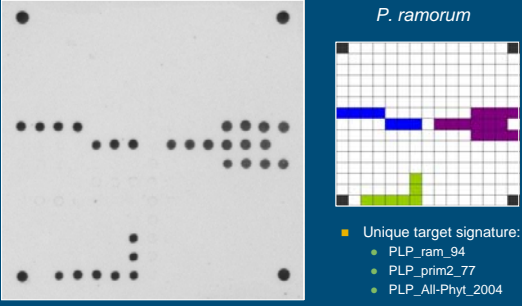


■ Specific ligation
■ No cross-hybridization
■ Unique target signature

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Individual target/mix of PLP's

P. ramorum



■ Unique target signature:
● PLP_ram_94
● PLP_prim2_77
● PLP_All-Phyt_2004

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Target signature for:

A: *P. alni*
B: *P. andina*
C: *P. brassicae*
D: *P. cinnamomi*
E: *P. cryptogea*
F: *P. fragariae*
G: *P. megasperma*
H: *P. primulae*



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DNA barcodes



DNA barcoding is a new technique that uses a short sequence of a standard region of the genomic DNA as a molecular diagnostic marker for species identification

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QBOL:

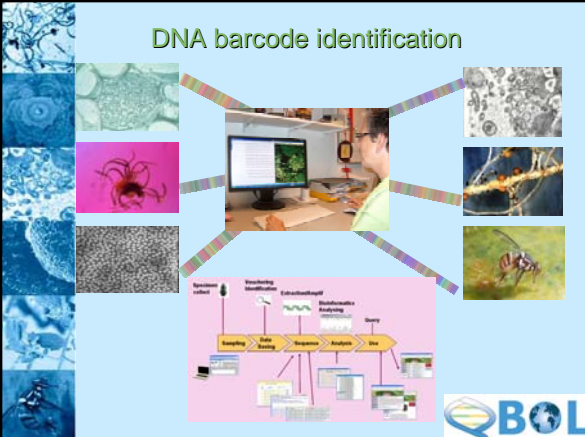
Development of a new diagnostic tool using DNA barcoding to identify quarantine organisms in support of plant health

www.qbol.org
peter.bonants@wur.nl




QBOL | Barcoding of Life

DNA barcode identification



Species identification
Identification of quarantine organisms
Identification of plant health

Sample → DNA Extraction → Sequencing → Analysis → Identification



DNA barcoders

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Espero que ustedes
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Muchas gracias

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